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 (30) Priority Data: 08/553,727 23 October 1995 (23.10.95) (71) Applicant: TULARIK, INC. [US/US]; Two Corpora South San Francisco, CA 94080 (US). (72) Inventors: BAICHWAL, Vijay, R.; Two Corpora South San Francisco, CA 94080 (US). HUANG, 	te Driv	Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.
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SCREENING ASSAYS

(57) Abstract

The invention relates to a human Receptor Interacting Protein (hRIP), nucleic acids which encode hRIP and methods of using the subject compositions; in particular, methods such as hRIP-based in vitro binding assays and phosphorylation assays for screening chemical libraries for lead compounds for pharmacological agents.

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RIP: Novel Human Protein Involved in Tumor Necrosis Factor Signal Transduction, and Screening Assays

INTRODUCTION

5 Field of the Invention

The field of this invention is a novel human kinase involved in tumor necrosis factor signal transduction and its use in drug screening.

Background

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Tumor necrosis factor (TNF) is an important cytokine involved in the signaling of a number of cellular responses including cytotoxicity, anti-viral activity, immun.-regulatory activities and the transcriptional regulation of a number of genes. The TNF receptors (TNF-R1 and TNF-R2) are members of the larger TNF receptor superfamily which also includes the Fas antigen, CD27, CD30, CD40, and the low affinity nerve growth factor receptor. Members of this family have been shown to participate in a variety of biological properties, including programmed cell death, antiviral activity and activation of the transcription factor NF- κ B in a wide variety of cell types.

Accordingly, it is desired to identify agents which specifically modulate transduction of TNF receptor family signalling. Unfortunately, the components of the signalling pathway remain largely unknown; hence, the reagents necessary for the development of high-throughput screening assays for such therapeutics are unavailable. Elucidation of TNF receptor family signal transduction pathways leading to NF-kB activation would provide valuable insight into mechanisms to alleviate inflammation. In particular, components of this pathway would provide valuable targets for automated, cost-effective, high throughput drug screening and hence would have immediate application in a broad range of domestic and international pharmaceutical and biotechnology drug development programs.

Relevant Literature

Stanger et al. (1995) Cell 81, 513-523 report the existence of a Receptor Interacting Protein (RIP) and its functional expression. VanArsdale and Ware (1994) J Immunology 153:3043-3050 describe proteins associated with TNF-R1. The cloning and amino acid sequencing of TNF-R1 is disclosed in Schall et al (1990) Cell 61, 361 and Loetscher et al (1990) Cell 61, 351; the identification of a "death domain" in TNF-R1 is disclosed in Tartaglia et al. (1993) Cell 74:845-853. The cloning and amino acid sequence of a TNF-R associated death domain protein (TRADD) is described by Hsu et al. (1995) Cell 81, 495-504. The cloning and amino acid sequence of the Fas antigen is disclosed in Itoh et al (1991)

Cell 66, 233-243. For a recent review, see Smith et al. (1994) Cell 76:959-962 and Vandenabelle et al. (1995) Trends Cell Biol. 5, 392-399.

SUMMARY OF THE INVENTION

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The invention provides methods and compositions relating to a human Receptor Interacting Protein (hRIP). The compositions include nucleic acids which encode hRIP, hRIP kinase domains, and recombinant proteins made from these nucleic acids. The invention also provides methods for screening chemical libraries for lead compounds for a pharmacological agent useful in the diagnosis or treatment of disease associated hRIP activity or hRIP-dependent signal transduction. In one embodiment, the methods involve incubating a mixture of hRIP, a natural intracellular hRIP substrate or binding target and a candidate pharmacological agent and determining if the presence of the agent modulates the ability of hRIP to selectively phosphorylate the substrate or bind the binding target. Specific agents provide lead compounds for pharmacological agents capable of disrupting hRIP function.

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DETAILED DESCRIPTION OF THE INVENTION

A human RIP-encoding nucleic acid sequence is set out in SEQ ID NO: 1. A human RIP kinase domain-encoding nucleic acid sequence is set out in SEQ ID NO: 1, nucleotides 1-900. A human RIP amino acid sequence is set out in SEQ ID NO: 2; and a hRIP kinase domain sequence is set out in SEQ ID NO: 2, residues 1-300.

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Natural nucleic acids encoding hRIP are readily isolated from cDNA libraries with PCR primers and hybridization probes containing portions of the nucleic acid sequence of SEQ ID NO:1. For example, we used low stringency hybridization at 42°C (hybridization buffer: 20% formamide, 10 % Denhardt, 0.5% SDS, 5X SSPE; with membrane washes at room temperature with 5X SSPE/0.5% SDS) with a 120 base oligonucleotide probe (SEQ ID NO: 1, nucleotides 1728-1847) to isolate a native human RIP cDNA from a library prepared from human umbilical vein endothelial cells. In addition, synthetic hRIP-encoding nucleic acids may be generated by automated synthesis.

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The subject nucleic acids are recombinant, meaning they comprise a sequence joined to a nucleotide other than that to which sequence is naturally joined and isolated from a natural environment. The nucleic acids may be part of hRIP-expression vectors and may be incorporated into cells for expression and screening, transgenic animals for functional studies

(e.g. the efficacy of candidate drugs for disease associated with expression of a hRIP), etc. These nucleic acids find a wide variety of applications including use as templates for transcription, hybridization probes, PCR primers, therapeutic nucleic acids, etc.; use in detecting the presence of hRIP genes and gene transcripts, in detecting or amplifying nucleic acids encoding additional hRIP homologs and structural analogs, and in gene therapy applications.

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The invention provides efficient methods of identifying pharmacological agents or lead compounds for agents active at the level of a hRIP modulatable cellular function, particularly hRIP mediated TNF receptor or Tumor necrosis factor receptor associated Factor -2 (TRAF2) or TRADD-induced signal transduction. For example, we have found that a binding complex comprising TNF R1, TRADD, and hRIP exists in TNF-stimulated cells. Generally, the screening methods involve assaying for compounds which interfere with a hRIP activity such as kinase activity or TRAF2 or TRADD binding. The methods are amenable to automated, cost-effective high throughput screening of chemical libraries for lead compounds. Identified reagents find use in the pharmaceutical industries for animal and human trials; for example, the reagents may be derivatized and rescreened in in vitro and in vivo assays to optimize activity and minimize toxicity for pharmaceutical development. Target therapeutic indications are limited only in that the target cellular function be subject to modulation, usually inhibition, by disruption of the formation of a complex comprising hRIP and one or more natural hRIP intracellular binding targets including substrates or otherwise modulating hRIP kinase activity. Target indications may include infection, genetic disease, cell growth and regulatory or immunolgic dysfunction, such as neoplasia, inflammation. hypersensitivity, etc.

A wide variety of assays for binding agents are provided including labeled in vitro kinase assays, protein-protein binding assays, immunoassays, cell based assays, etc. The hRIP compositions used in the methods are recombinantly produced from nucleic acids having the disclosed hRIP nucleotide sequences. The hRIP may be part of a fusion product with another peptide or polypeptide, e.g. a polypeptide that is capable of providing or enhancing protein-protein binding, stability under assay conditions (e.g. a tag for detection or anchoring), etc.

The assay mixtures comprise one or more natural intracellular hRIP binding targets including substrates, such as TRADD, TRAF2, or, in the case of an autophosphorylation

assay, the hRIP itself can function as the binding target. In one embodiment, the mixture comprises a complex of hRIP, TRADD and TNFR1. A hRIP derived pseudosubstrate may be used or modified (e.g. A to S/T substitutions) to generate effective substrates for use in the subject kinase assays as can synthetic peptides or other protein substrates. Generally, hRIP-specificity of the binding agent is shown by kinase activity (i.e. the agent demonstrates activity of an hRIP substrate, agonist, antagonist, etc.) or binding equilibrium constants (usually at least about 10⁶ M⁻¹, preferably at least about 10⁸ M⁻¹, more preferably at least about 10⁹ M⁻¹. A wide variety of cell-based and cell-free assays may be used to demonstrate hRIP-specific binding; preferred are rapid in vitro, cell-free assays such as mediating or inhibiting hRIP-protein (e.g. hRIP-TRADD) binding, phosphorylation assays, immunoassays, etc.

The assay mixture also comprises a candidate pharmacological agent. Candidate agents encompass numerous chemical classes, though typically they are organic compounds; preferably small organic compounds and are obtained from a wide variety of sources including libraries of synthetic or natural compounds. A variety of other reagents may also be included in the mixture. These include reagents like salts, buffers, neutral proteins, e.g. albumin, detergents, etc. which may be used to facilitate optimal binding and/or reduce non-specific or background interactions, etc. Also, reagents that otherwise improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, antimicrobial agents, etc. may be used.

In a preferred in vitro, binding assay, a mixture of at least the kinase domain of hRIP, one or more binding targets or substrates and the candidate agent is incubated under conditions whereby, but for the presence of the candidate pharmacological agent, the hRIP specifically binds the cellular binding target at a first binding affinity or phosphoylates the substrate at a first rate. After incubation, a second binding affinity or rate is detected.

Detection may be effected in any convenient way. For cell-free binding assays, one of the components usually comprises or is coupled to a label. The label may provide for direct detection as radioactivity, luminescence, optical or electron density, etc. or indirect detection such as an epitope tag, an enzyme, etc. A variety of methods may be used to detect the label depending on the nature of the label and other assay components. For example, the label may be detected bound to the solid substrate or a portion of the bound complex containing the label may be separated from the solid substrate, and thereafter the label detected.

The following examples are offered by way of illustration and not by way of limitation.

EXAMPLES

- 5 1. Protocol for hRIP autophosphorylation assay.
 - A. Reagents:

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- Neutralite Avidin: 20 µg/ml in PBS.
- -hRIP: 10^8 10^5 M biotinylated hRIP kinase domain, residues 1-300 at 20 μ g/ml in PBS.
 - Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.
- Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.
- -[32 P]γ-ATP 10x stock: 2 x 10⁻⁵ M cold ATP with 100 μCi [32 P]γ-ATP. Place in the 4°C microfridge during screening.
- Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVo₃ (Sigma # S-6508) in 10 ml PBS.
 - B. Preparation of assay plates:
 - Coat with 120 µl of stock Neutralite avidin per well overnight at 4°C.
 - Wash 2 times with 200 µl PBS.
 - Block with 150 µl of blocking buffer.
 - Wash 2 times with 200 µl PBS.
 - C. Assay:
- 25 Add 40 μl assay buffer/well.
 - Add 40 µl biotinylated hRIP (0.1-10 pmoles/40 ul in assay buffer)
 - Add 10 µl compound or extract.
 - Add 10 μl [32P]γ-ATP 10x stock.
 - Shake at 30°C for 15 minutes.
- Incubate additional 45 minutes at 30°C.
 - Stop the reaction by washing 4 times with 200 µl PBS.
 - Add 150 µl scintillation cocktail.

- Count in Topcount.
- D. Controls for all assays (located on each plate):
 - a. Non-specific binding (no RIP added)
 - b. cold ATP to achieve 80% inhibition.

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- 2. Protocol for hRIP substrate phosphorylation assay.
- A. Reagents:
 - Neutralite Avidin: 20 µg/ml in PBS.
 - -<u>hRIP</u>: 10⁻⁸ 10⁻⁵ M hRIP at 20 μg/ml in PBS.

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- Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.
- Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.
- -[32 P]γ-ATP 10x stock: 2 x 10⁻⁵ M cold ATP with 100 μCi [32 P]γ-ATP. Place in the 4°C microfridge during screening.
- Substrate: 2 x 10⁻⁶ M biotinylated synthetic peptide kinase substrate at 20 μg/ml in PBS.
 - Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVo₃ (Sigma # S-6508) in 10 ml PBS.
 - B. Preparation of assay plates:
 - Coat with 120 µl of stock Neutralite avidin per well overnight at 4°C.
 - Wash 2 times with 200 µl PBS.
 - Block with 150 µl of blocking buffer.
- Wash 2 times with 200 μl PBS.
 - C. Assay:
 - Add 40 µl assay buffer/well.
 - Add 40 µl hRIP (0.1-10 pmoles/40 ul in assay buffer)
 - Add 10 µl compound or extract.
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- Shake at 30°C for 15 minutes.
- Add 10 μl [³²P]γ-ATP 10x stock.
- Add 10 µl substrate.

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- Shake at 30°C for 15 minutes.
- Incubate additional 45 minutes at 30°C.
- Stop the reaction by washing 4 times with 200 µl PBS.
- Add 150 µl scintillation cocktail.
- 5 Count in Topcount.
 - D. Controls for all assays (located on each plate):
 - a. Non-specific binding (no RIP added)
 - b. cold ATP to achieve 80% inhibition.
- 10 3. Protocol for hRIP TRADD binding assay.
 - A. Reagents:

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- Anti-myc antibody: 20 µg/ml in PBS.
- Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.
- Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM β-mercaptoethanol, 1 mg/ml BSA, cocktail of protease inhibitors.
- ²³P hRIP 10x stock: 10⁻⁸ 10⁻⁶M "cold" hRIP (full length) supplemented with 200,000-250,000 cpm of labeled hRIP (HMK-tagged) (Beckman counter). Place in the 4°C microfridge during screening.
- Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVo₃ (Sigma # S-6508) in 10 ml PBS.
 - TRADD: 10⁻⁸ 10⁻⁵ M myc eptitope-tagged TRADD in PBS.
 - B. Preparation of assay plates:
 - Coat with 120 µl of stock anti-myc antibody per well overnight at 4°C.
 - Wash 2X with 200 µl PBS.
 - Block with 150 µl of blocking buffer.
 - Wash 2X with 200 µl PBS.
 - C. Assay:
- Add 40 μl assay buffer/well.
 - Add 10 µl compound or extract.
 - Add 10 μ l ³³P-RIP (20,000-25,000 cpm/0.1-10 pmoles/well =10⁻⁹- 10⁻⁷ M final

concentration).

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- Shake at 25°C for 15 minutes.
- Incubate additional 45 minutes at 25°C.
- Add 40 µl eptitope-tagged TRADD (0.1-10 pmoles/40 ul in assay buffer)
- Incubate 1 hour at room temperature.
 - Stop the reaction by washing 4 times with 200 µl PBS.
 - Add 150 µl scintillation cocktail.
 - Count in Topcount.
 - D. Controls for all assays (located on each plate):
 - a. Non-specific binding (no hRIP added)
 - b. Soluble (non-tagged TRADD) to achieve 80% inhibition.
 - 4. Protocol for hRIP TRAF2 binding assay.
 - A. Reagents:
- 15 Anti-myc antibody: 20 μg/ml in PBS.
 - Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.
 - Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM β-mercaptoethanol, 1 mg/ml BSA, cocktail of protease inhibitors.
 - ³³P hRIP 10x stock: 10⁻⁸ 10⁻⁶ M "cold" hRIP kinase domain, residues 1-300, supplemented with 200,000-250,000 cpm of labeled hRIP kinase domain (HMK-tagged) (Beckman counter). Place in the 4°C microfridge during screening.
 - Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVo₃ (Sigma # S-6508) in 10 ml PBS.
 - TRAF2: 10⁻⁸ 10⁻⁵ M myc eptitope-tagged TRAF2 in PBS.
 - B. Preparation of assay plates:
 - Coat with 120 µl of stock anti-myc antibody per well overnight at 4°C.
 - Wash 2X with 200 µl PBS.
 - Block with 150 µl of blocking buffer.
 - Wash 2X with 200 µl PBS.
 - C. Assay:

- Add 40 µl assay buffer/well.
- Add 10 µl compound or extract.
- Add 10 μ l ³³P-RIP kinase domain (20,000-25,000 cpm/0.1-10 pmoles/well =10⁻⁹- 10⁻⁷ M final concentration).
 - Shake at 25°C for 15 minutes.
 - Incubate additional 45 minutes at 25°C.
 - Add 40 µl eptitope-tagged TRAF2 (0.1-10 pmoles/40 ul in assay buffer)
 - Incubate 1 hour at room temperature.
 - Stop the reaction by washing 4 times with 200 μl PBS.
 - Add 150 µl scintillation cocktail.
 - Count in Topcount.
- D. Controls for all assays (located on each plate):
 - a. Non-specific binding (no hRIP kinase domain added)
 - b. Soluble (non-tagged TRAF2) to achieve 80% inhibition.

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All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference. Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

SEQUENCE LISTING

	SEQUENCE LISTING
	(1) GENERAL INFORMATION:
	(i) APPLICANT: BAICHWAL, VIJAY R
	HUANG, JIANING
· 5	HSU, HAILING
	GOEDDEL, DAVID V
	(ii) TITLE OF INVENTION: RIP: NOVEL HUMAN PROTEIN INVOLVED IN
	TUMOR NECROSIS FACTOR SIGNAL TRANSDUCTION, AND SCREENING
	ASSAYS
10	(iii) NUMBER OF SEQUENCES: 2
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	(C) CITY: SAN FRANCISCO
15	(D) STATE: CALIFORNIA
	(E) COUNTRY: USA
	(F) ZIP: 94111-4187
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20	(B) COMPUTER: IBM PC compatible
	(C) OPERATING SYSTEM: PC-DOS/MS-DOS
	(D) SOFTWARE: PatentIn Release #1.0, Version #1.30
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25	(B) FILING DATE:
	(C) CLASSIFICATION:
	(viii) ATTORNEY/AGENT INFORMATION:
	(A) NAME: BREZNER, DAVID J
20	(B) REGISTRATION NUMBER: 24,774
30	(C) REFERENCE/DOCKET NUMBER: T95-006/PCT
	(ix) TELECOMMUNICATION INFORMATION:
	(A) TELEPHONE: (415) 781-1989
	(B) TELEFAX: (415) 398-3249
25	
35	(2) INFORMATION FOR SEQ ID NO:1:
	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 2016 base pairs
	(B) TYPE: nucleic acid
40	(C) STRANDEDNESS: double
70	(D) TOPOLOGY: linear
	(ii) MOLECULE TYPE: cDNA
	(ix) FEATURE:
	(A) NAME/KEY: CDS
45	(B) LOCATION: 12013
15	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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	CTT	CAA	CTT	GAT	TGT	GTG	GCA	GTA	CCT	TCA	AGC	CGG	TCA	AAT	TCA	GCC	1008
	Leu	Gln	Leu	Asp	Cys	Val	Ala	Val	Pro	Ser	Ser	Arg	Ser	Asn	Ser	Ala	
					325					330					335		
																GGT	1056
20	Thr	Glu	Gln		Gly	Ser	Leu	His		Ser	Gln	Gly	Leu			Gly	
				340		-			345	m oo	ama	010	010	350		CAA	1104
																GAA Glu	1104
	PIO	Val	355		361	لإلا	rne	360	110	Jer	Dea	014	365				
25	GAG	AAT			AGC	CTG	CAG		AAA	CTC	CAA	GAC			AAC	TAC	1152
																Tyr	
		370					375					380					
	CAT	CTI	TAT	GGC	AGC	CGC	ATG	GAC	AGG	CAG	ACG	AAA	CAG	CAG	CCC	AGA	1200
	His	Lev	Туг	Gly	Ser	Arg	Met	Asp	Arg	Gln	Thr	Lys	Glr	Glr	Pro	Arg	
30	385					390					395					400	
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	Gln	Asr	ı Val	Ala		_	Arg	Glu	Glu			Arg	Arc	Arg		L Ser -	
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35																r ACA n Thr	
22	urs	, MSI) PIC	420		ı Gii	GII	. MI	42		GIC	. voi	1 2110	430			
	GAC	GG/	A AA			r GTT	rat 1	TC			A GCC	AG	r car			r GCA	. 1344
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40	GT	G CA	C CA	G CC	C TC	A GG	CTO	: AC	C AG	C CA	A CC	r CA	A GT	A CT	G TA	T CAG	1392
	Va.	l Hi	s Gl	n Pro	o Se	r Gl	y Lev	Th:	r Se	r Gl	n Pro	o G1:	n Va	l Le	и Ту	r Gln	1
		45					455					46					
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	As	n As	n Gl	y Le	u Ty	r Se	r Se	r Hi	s Gl	y Ph			r Ar	g Pr	o Le	u Asp	
45	46	5				47	0				47	5				480)

•	CCA	GGA	ACA	GCA	GGT	CCC	AGA	GTT	TGG	TAC	AGG	CCA	ATT	CCA	AGT	CAT	1488
	Pro	Gly	Thr	Ala	Gly	Pro	Arg	Val	Trp	Tyr	Arg	Pro	Ile	Pro	Ser	His	
					485					490					495		
	ATG	CCT	AGT	CTG	CAT	AAT	ATC	CCA	GTG	CCT	GAG	ACC	AAC	TAT	CTA	GGA	1536
5	Met	Pro	Ser	Leu	His	Asn	Ile	Pro	Val	Pro	Glu	Thr	Asn	Tyr	Leu	Gly	
				500					505					510			
	AAT	ACA	ccc	ACC	ATG	CCA	TTC	AGC	TCC	TTG	CCA	CCA	ACA	GAT	GAA	TCT	1584
	Asn	Thr	Pro	Thr	Met	Pro	Phe	Ser	Ser	Leu	Pro	Pro	Thr	Asp	Glu	Ser	
			515					520					525				
10	ATA	AAA	TAT	ACC	ATA	TAC	AAT	AGT	ACT	GGC	ATT	CAG	ATT	GGA	GCC	TAC	1632
	Ile	Lys	Tyr	Thr	Ile	Tyr	Asn	Ser	Thr	Gly	Ile	Gln	Ile	Gly	Ala	Tyr	
		530					535					540					
	AAT	TAT	ATG	GAG	ATT	GGT	GGG	ACG	AGT	TCA	TCA	CTA	CTA	GAC	AGC	ACA	1680
	Asn	Tyr	Met	Glu	Ile	Gly	Gly	Thr	Ser	Ser	Ser	Leu	Leu	Asp	Ser	Thr	•
15	545					550					555					560	
	AAT	ACG	AAC	TTC	AAA	GAA	GAG	CCA	GCT	GCT	AAG	TAC	CAA	GCT	ATC	TTT	1728
	Asn	Thr	Asn	Phe	Lys	Glu	Glu	Pro	Ala	Ala	Lys	Tyr	Gln	Ala	Ile	Phe	
					565					570					575		
					AGT												1776
20	Asp	Asn	Thr	Thr	Ser	Leu	Thr	Asp	Lys	His	Leu	Asp	Pro	Ile	Arg	Glu	
				580					585					590			
					CAC												1824
	Asn	Leu	Gly	Lys	His	Trp	Lys	Asn	Cys	Ala	Arg	Lys	Leu	Gly	Phe	Thr	
			595					600				,	605				
25																CTG	1872
	Gln	Ser	Gln	Ile	Asp	Glu	Ile	Asp	His	Asp	Туг			Asp	Gly	Leu	
		610					615					620					
																GGC	1920
	Lys	Glu	Lys	Val	Tyr			Leu	Gln	Lys			Met	Arg	Glu	Gly	
30	625					630					635					640	
																TGT	1968
	Ile	Lys	Gly	Ala			Gly	Lys	Leu			Ala	Lev	His		Cys	
					645					650					655		
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35	Ser	Arg	, Ile		Leu	Leu	Ser	Ser			Туг	· Val	Ser			1	
				660)				665	•				670	J		
	TAI	4															2016

(2) INFORMATION FOR SEQ ID NO:2:

- 40 (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 671 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
- 45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

	Met	Gln	Pro	Asp	Met	Ser	Leu	Asn	Val	Ile	гàг	Met	Lys	Ser	Ser	Asp
	1				5					10					15	
	Phe	Leu	Glu	Ser	Ala	Glu	Leu	qaA	Ser	Gly	Gly	Phe	Gly	Lys	Val	Ser
				20					25	_				30		
5	T.011	Cvs	Phe		Ατσ	Thr	Gln	Glv		Met	Tle	Met	Ive		Val	ጥህን
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	_		35	_	_		_,					_				
	Lys	_	Pro	Asn	Cys	Ile		His	Asn	GLu	Ala		Leu	Glu	Glu	Ala
		50					55					60				
	Lys	Met	Met	Asn	Arg	Leu	Arg	His	Ser	Arg	Val	Val	Lys	Leu	Leu	Gly
10	65					70					75					80
	Val	Ile	Ile	Glu	Glu	Gly	Lys	Tyr	Ser	Leu	Val	Met	Glu	Tyr	Met	Glu
					85					90					95	
	Lvs	Glv	Asn	Leu	Met	His	Val	Leu	Lvs	Ala	Glu	Met	Ser	Thr	Pro	Leu
	•			100					105					110		
15	Com	37-1	Lys		3 ~~ ~	710	T10	T		T10	T10	C111	G1 ₁₄			Th
13	Set	Val	-	GIY	ALY	116	116		GIU	116	116	GIU		Mec	Cys	TAT
	_		115	_		•		120	_	_	_	_	125		_	
	Leu		Gly	Lys	GIA	Val		His	Lys	qaA	Leu		Pro	GIu	Asn	Ile
		130					135					140				
	Leu	Val	Asp	Asn	Asp	Phe	His	Ile	Lys	Ile	Ala	Asp	Leu	Gly	Leu	Ala
20	145					150					155					160
	Ser	Phe	Lys	Met	Trp	Ser	Lys	Leu	Asn	Asn	Glu	Glu	His	Asn	Glu	Leu
					165					170					175	
	Arg	Glu	Val	Asp	Gly	Thr	Ala	Lys	Lys	Asn	Gly	Gly	Thr	Leu	Tyr	Tyr
				180					185					190		
25	Met	Ala	Pro	Glu	His	Leu	Asn	Asp			Ala	Lvs	Pro	Thr	Glu	Lvs
			195					200				-1-	205			-1-
	C.~					Dho	31 n			T 011	П -	21.			71 5) an
	Set		Val	ıyı	Ser	PHE			vaı	Leu	ııp			FIIE	Ala	ASII
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30	225	5				230					235					240
	Ile	Lys	Ser	Gly	Asn	Arg	Pro	Asp	Val	. Asp	Asp	Ile	Thr	Glu	Tyr	Cys
					245					250)				255	
	Pro	Arg	, Glu	Ile	: Ile	Ser	Leu	Met	Lys	Lev	Cys	Trp	Glu	Ala	Asn	Pro
				260)				265	5				270		
35	Glu	ı Ala	a Arq	Pro	Thr	Phe	Pro	Gly	, Ile	e Glu	ı Glu	Lys	Phe	Arg	Pro	Phe
			275					280				•	285			
	The sa	r T.o.			La	. G1:	G31			(1)	. G1v	Acr			50*	Leu
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40	30					310					315					320
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					329	5				33	0				335	i
	Th	r Gl	u Glr	n Pro	o Gly	/ Sei	. Le	ı His	s Se	r Se	r Glr	ı Gly	/ Leu	ı Gly	/ Met	Gly
				340	0		-		34	5				350)	
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	Pro	Val	Glu	Glu	Ser	Trp	Phe	Ala	Pro	Ser	Leu	Glu	His	Pro	Gln	Glu
			355					360					365			
	Glu	Asn	Glu	Pro	Ser	Leu	Gln	Ser	Lys	Leu	Gln		Glu	Ala	Asn	Tyr
_		370		_			375	_	_			380			_	_
5		Leu	Tyr	Gly	Ser		Met	Asp	Arg	Gln		Lys	Gln	GIn	Pro	
	385				_	390		~1	-1	~1	395	•	•		··- 1	400
	Gln	Asn	Val	Ala		Asn	Arg	Giu	GIU		Arg	Arg	Arg	Arg	Val	ser
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10	HIS	Asp	Pro		Ald	GIII	GIII	AIG		ıyı	GIU	POII	FILE	430	Asn	1111
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	Val	His		Pro	Ser	Glv	Leu		Ser	Gln	Pro	Gln		Leu	Tyr	Gln
		450					455					460			-	
15	Asn		Gly	Leu	Tyr	Ser	Ser	His	Gly	Phe	Gly	Thr	Arg	Pro	Leu	Asp
	465					470					475					480
	Pro	Gly	Thr	Ala	Gly	Pro	Arg	Val	Trp	Tyr	Arg	Pro	Ile	Pro	Ser	His
					485					490					495	
	Met	Pro	Ser	Leu	His	Asn	Ile	Pro	Val	Pro	Glu	Thr	Asn	Tyr	Leu	Gly
20				500					505					510		
	Asn	Thr	Pro	Thr	Met	Pro	Phe		Ser	Leu	Pro	Pro		Asp	Glu	Ser
			515					520			_	_	525			_
	Ile		Tyr	Thr	Ile	Tyr		Ser	Thr	Gly	Ile		Ile	GIĀ	Ala	TYT
26		530					535					540	•	•	a	mb
25			Met	Glu	TIE		GTA	Thr	Ser	ser	5er 555	Leu	Leu	Asp	Ser	560
	545		Acn	Phe	Larg	550	Glu	Pro	Δla	λla		ጥህን	Gln	Ala	Ile	
	ASII		ASII	rne	565		910	110	77.4	570		-,-	01		575	
	Asp	Asn	Thr	Thr			Thr	Asp	Lys			Asp	Pro	Ile	Arg	Glu
30				580				•	585			_		590		
	Asn	Leu	Gly	Lys	His	Trp	Lys	Asn	Cys	Ala	Arg	Lys	Leu	Gly	Phe	Thr
			5 95					600					605			
	Gln	Ser	Gln	Ile	Asp	Glu	Ile	Asp	His	Asp	Туг	Glu	Arg	Asp	Gly	Leu
		610	ŀ				615					620				
35	Lys	Glu	Lys	Val	Tyr	Gln	Met	Leu	Gln	Lys	Trp	Val	Met	Arg	Glu	Gly
	625					630					635					640
	Ile	Lys	Gly	Ala	Thr	Val	. Gly	Lys	Leu	Ala	Glr	Ala	Leu	His	Gln	Cys
					645			,		650					655	
40	Ser	Arg	, Ile			Let	. Ser	Sex			ту:	Val	Ser		Asn	L
40				660)				665	•				670)	

WHAT IS CLAIMED IS:

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1. An isolated, recombinant nucleic acid encoding a human Receptor Interacting Protein (hRIP) kinase domain.

- 5 2. An isolated, recombinant nucleic acid encoding a human Receptor Interacting Protein (hRIP) comprising SEQ ID NO: 1.
 - 3. A method of making a human Receptor Interacting Protein (hRIP) kinase domain containing protein, said method comprising the steps of translating a nucleic acid according to claim 1 to form a translation product and isolating said translation product.
 - 4. A method of identifying lead compounds for a pharmacological agent useful in the diagnosis or treatment of disease, said method comprising the steps of:

making a protein according to the method of claim 3,

forming a mixture comprising:

said protein,

a natural intracellular hRIP binding target, wherein said binding target is capable of specifically binding said protein, and

a candidate pharmacological agent;

incubating said mixture under conditions whereby, but for the presence of said candidate pharmacological agent, said protein selectively binds said binding target at a first binding affinity;

detecting a second binding affinity of said protein to said binding target,

- wherein a difference between said first and second binding affinity indicates that said candidate pharmacological agent is a lead compound for a pharmacological agent capable of modulating hRIP-dependent signal transduction.
- 5. A method according to claim 4, wherein said hRIP binding target comprises a Tumor necrosis factor receptor Associated Factor -2 (TRAF2) or a Tumor necrosis factor Receptor-1 Associated Death Domain protein (TRADD).

6. A method of identifying lead compounds for a pharmacological agent useful in the diagnosis or treatment of disease, said method comprising the steps of:

making a protein according to the method of claim 3,

forming a mixture comprising:

said protein,

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an hRIP substrate, wherein said hRIP kinase domain of said protein is capable of specifically phosphorylating said substrate, and

a candidate pharmacological agent;

incubating said mixture under conditions whereby, but for the presence of said candidate pharmacological agent, said hRIP kinase domain selectively phosphorylates said substrate at a first rate;

detecting a second rate of phosphorylation of said substrate by said hRIP kinase domain,

wherein a difference between said first and second rate indicates that said candidate pharmacological agent is a lead compound for a pharmacological agent capable of modulating hRIP kinase activity.

7. A method according to claim 6 wherein said hRIP substrate is hRIP.

International application No. PCT/US96/16778

A. CLA	SSIFICATION OF SUBJECT MATTER	-							
(-/	:Please See Extra Sheet. : 536/23.5; 435/ 69.1, 69.5, 252.3, 320.1; 530/350, 3	251							
According to	o International Patent Classification (IPC) or to both n	national classification and IPC							
B. FIEL	DS SEARCHED								
Minimum d	ocumentation searched (classification system followed	by classification symbols)							
U.S . :	U.S. : 536/23.5; 435/ 69.1, 69.5, 252.3, 320.1; 530/350, 351								
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched									
Electronic d	ata base consulted during the international search (nar	ne of data base and, where practicable,	search terms used)						
C. DOC	UMENTS CONSIDERED TO BE RELEVANT								
Category*	Citation of document, with indication, where app	Relevant to claim No.							
x	STANGER et al. RIP: A Novel Pro	tein Containing a Death	1-3						
	Domain That Interacts with Fas/AF	-							
Υ	Causes Cell Death. Cell. 19 May 1995, Vol. 81, pages 513- 2 523, see Figs. 2-3, and sequence alignment,								
Y, P	WO 96/25941 A1 (YEDA RESEAR) LTD.) 29 August 1996 (29/08/96), claims.	1-3							
A	HSU et al. The TNF Receptor 1-As Signals Cell Death and NF-kB Active Vol. 81, pages 495-504, see all.		1-3						
X Furt	her documents are listed in the continuation of Box C	. See patent family annex.							
·	pocial estegories of cited documents:	"T" Inter document published after the int date and not in conflict with the applic	ention but cited to understand the						
	ocument defining the general state of the art which is not considered be of particular relevance	"X" document of particular relevance; the							
	rijer document published on or after the international filing date ocument which may throw doubts on priority claim(s) or which is	"X" document of particular relevance; it considered novel or cannot be considered novel or cannot be considered novel or cannot in taken alone							
\ ci	ted to establish the publication date of another citation or other secial reason (as specified)	"Y" document of particular relevance; the							
•0• de	ocument referring to an oral disclosure, use, exhibition or other tons	considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.							
	ocument published prior to the international filing date but later than se priority date claimed	*&* document member of the same patent family							
Date of the	actual completion of the international search	Date of mailing of the international search report							
15 JANU	JARY 1997	2 8FEB 1997							
Name and Commissi Box PCT	mailing address of the ISA/US oner of Patents and Trademarks	Authorized officer Tiu-To	^						
Washingto	oa, D.C. 20231	GARNETTE D. DRAPER							
Facsimile	No. (703) 305-3230	Telephone No. (703) 308-0196							

International application No.
PCT/US96/16778

C (Continua	tion). DOCUMENTS CONSIDERED TO BE RELEVANT										
Category*	Citation of document, with indication, where appropriate, of the relev	tation of document, with indication, where appropriate, of the relevant passages									
А, Р	BAKER et al. Transducers of Life and Death: TNF Re Superfamily and Associated Proteins. Oncogene, 04 Ja Vol. 12, pages 1-9, see all	1-3									
į											
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International application No. PCT/US96/16778

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
Picase See Extra Sheet.
As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment
of any additional fee. 3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. X No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-3
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

International application No. PCT/US96/16778

A. CLASSIFICATION OF SUBJECT MATTER: IPC (6):

C07H 21/04; C12P 21/06, 21/02; C12N 1/20, 15/00; C07K 1/00, 14/52

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claims 1-3, drawn to nucleic acids that encode for human Receptor Interacting Proteins (hRIP) and methods of making the encoded proteins.

Group II, claims 4-7, drawn to methods of identifying lead compounds.

The inventions listed as Groups do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: The special technical feature of Group I is directed to nucleic acids that encode for hRIP and to methods of making hRIP; whereas the special technical feature of Group II is directed to methods of identifying lead compounds. The methods of these two groups do not share a special technical and unifying feature, because each of these methods require the utilization of different process/method steps, different elements/agents, and their are different starting material and the final outcomes are also different. Furthermore, these methods and their steps and elements are not required one for the other.